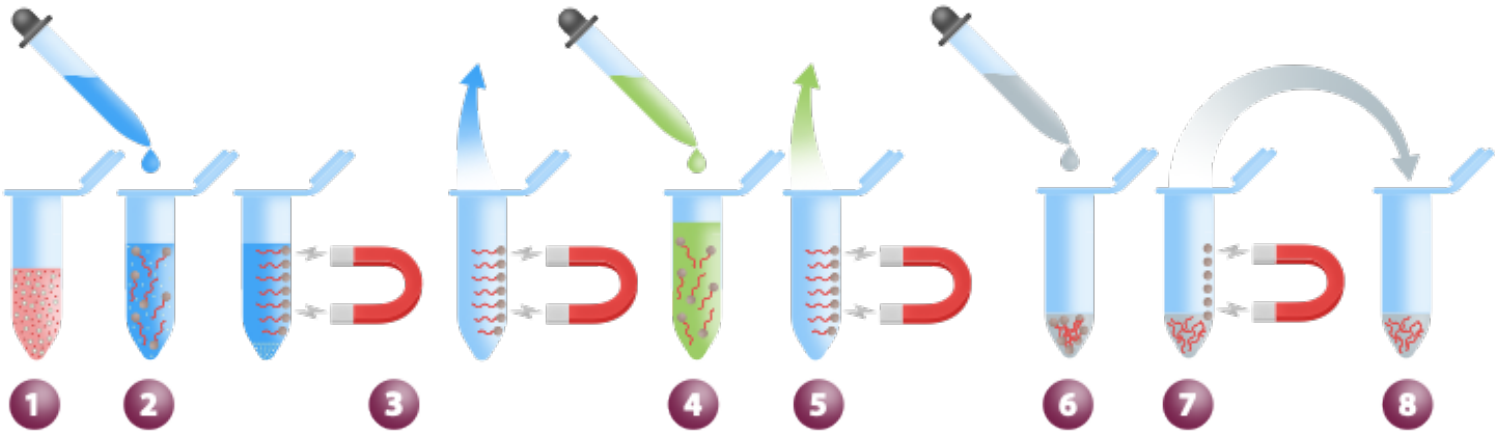
magnetiC

Viral RNA Purification

Magnetic Bead-Based RNA Isolation from Complex Body Fluids



1. In 1.5 mL microfuge tube, add 20 µL of Proteinase K and 100 µL of sample (serum/plasma, saliva, sputum, cell/mucus-laden swab samples or culture supernatants)
   Optional: If extracting/purifying low RNA concentrations, also add 2.5 µL *of* **Carrier RNA** *to sample*
   Note: Sputum/Swab samples may need to be vortexed, then centrifuged to remove debris
2. Add 400 µL of **Viral Lysis/Binding Buffer** and mix well by pipetting up-down 10-15x. In lieu of manual mixing, pulse vortex for 15 seconds. Incubate 5 mins to allow for lysis and RNA binding.
   For vortexed samples, perform rapid 2-5 second spin following incubation
3. Place tube on magnetic rack for 1-2 mins to capture RNA-bead complex, then discard supernatant
4. Remove tube from magnetic rack and resuspend RNA/bead complex in 600 µL of **Wash Buffer #1**
5. Return to magnetic rack for 5 mins, then discard supernatant. Repeat wash with 600 µL **Wash Buffer #2**, return to magnetic rack for 1-2 mins, then discard supernatant and leave to dry for 1 min
6. Remove tube from magnetic rack and resuspend RNA-bead complex in 50 µL of **Elution Buffer**. Mix well by pipetting up-down 10-15x to elute RNA from beads and let stand for 1-2 mins
7. Place tube on magnetic rack to separate beads (~1-2 mins)
8. Transfer clean RNA solution (supernatant) to clean tube

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